

# Antimicrobial properties of sulphur-enriched, hydrophilic MoS<sub>2</sub> nano/microparticles and heterostructured Pd/MoS<sub>2</sub>/Ti coatings

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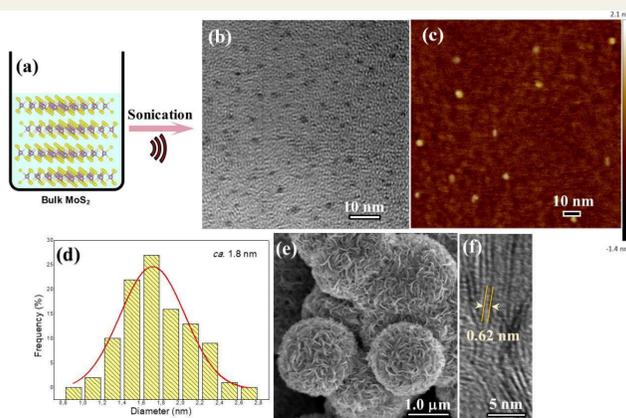


## Introduction

Recently, graphene-like two-dimensional molybdenum disulphide-based nanomaterials, especially their single or few-layered forms, usually named nanosheets (MoS<sub>2</sub>-ns) or nanoknives, have received considerable attention as a promising antimicrobial agent. However, most previous studies indicate that without functionalization with other antimicrobial agents such as Ag, Ti<sub>3</sub>C<sub>2</sub>MXene, graphene oxide (GO) [1, 2], etc., the antimicrobial efficiency of MoS<sub>2</sub> is low and needs further improvements. In this study, the MoS<sub>2</sub>-based nano/microparticles and coatings were synthesized through a simple, one-step hydrothermal approach without any other additives. The fabricated materials exhibited relatively small ( $\Delta\theta = 18.7 \pm 2.5^\circ$ ) contact angle, resulting in their prominent hydrophilic properties, possibly caused due to sulphur-enriched MoS<sub>2</sub> composite as evidenced by TG/DTA-MS analysis. Such nanostructures can exhibit a better adhesion of biomolecules, thus facilitating the interaction between them, as confirmed by highly effective antimicrobial action (Fig. 1). The present study examines the antimicrobial properties of hydrophilic, sulphur-enriched MoS<sub>2</sub> nano/microparticles as well as MoS<sub>2</sub>-based coatings against various humans' pathogenic bacteria such as *S. enterica*, *P. aeruginosa*, *E. coli*, *S. aureus* (MRSA), *M. luteus*, and two *Candida* fungi, in particular *C. parapsilosis*, *C. krusei*. The MoS<sub>2</sub>-ns (40  $\mu\text{g mL}^{-1}$ ) showed over 90 % killing efficiency against *S. aureus* MRSA bacteria and two *Candida* fungi within 24 h of exposure. Surprisingly, the petal-like MoS<sub>2</sub> microstructures and heterostructured MoS<sub>2</sub>/Ti and Pd/MoS<sub>2</sub>/Ti coatings also possess high antimicrobial potency and could be considered a promising antimicrobial agent and thus deserve further studies. The MoS<sub>2</sub>-induced intracellular reactive oxygen species (ROS) production was evidenced by measuring the standard DCF dye fluorescence.

## Characterization of the MoS<sub>2</sub> NPs, petal-like MoS<sub>2</sub> spheres

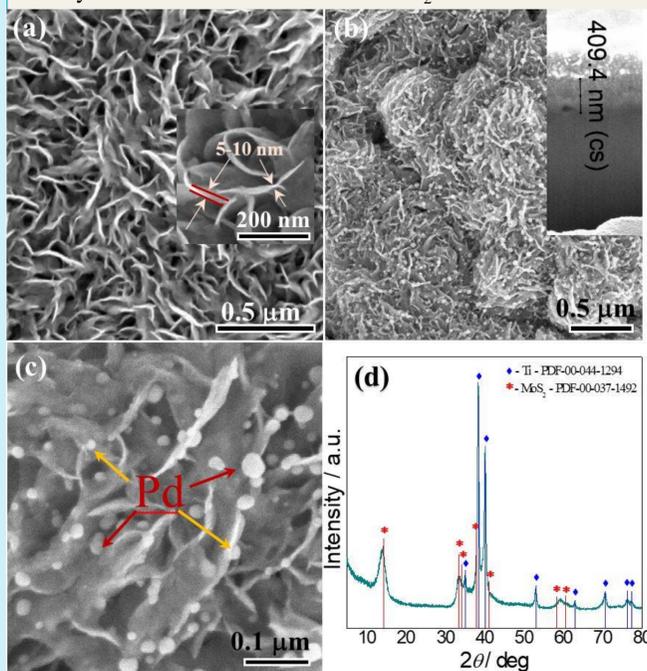
The structural analysis of the as-grown product was performed using TEM and AFM techniques. Morphology investigations of MoS<sub>2</sub>-ns revealed that these NPs have a narrow size distribution between 0.8 and 2.8 nm with an average diameter of 1.8 nm (Fig. 1 b, d). TEM results match well those determined by atomic force microscopy, which showed that these NPs were exfoliated to a few-layered material with a narrow size distribution (Fig. 1 c). The spherical structure of black MoS<sub>2</sub> powder with petal-like surface design (as presented in Fig. 1 e) was synthesized by a hydrothermal approach. SEM image depicted that bulk MoS<sub>2</sub> are mainly spherical with average diameters of 1.5–2.5  $\mu\text{m}$  and their surface is uniformly covered with differently oriented leaflets. In order to prove their lamellar structure, the individual leaflet edges were assessed by TEM. Figure 1 f indicate lamellar morphology with dominant interlayer spacing approximately equal to 0.62 Å attributable to (002) plane of MoS<sub>2</sub>.



**Fig. 1.** TEM (b) and AFM (c) images of nanosheets obtained by ultrasonic exfoliation procedure (a) of hydrothermally synthesized MoS<sub>2</sub> powder. Histogram of size distribution acquired from multiple TEM images is shown in (d). SEM image of hydrothermally produced MoS<sub>2</sub> black powder resembles petal-like spheres (e). In (f), HR-TEM image of nanosheet edges.

## Morphology and phase investigation of nanoplatelet MoS<sub>2</sub>/Ti and Pd/MoS<sub>2</sub>/Ti coatings

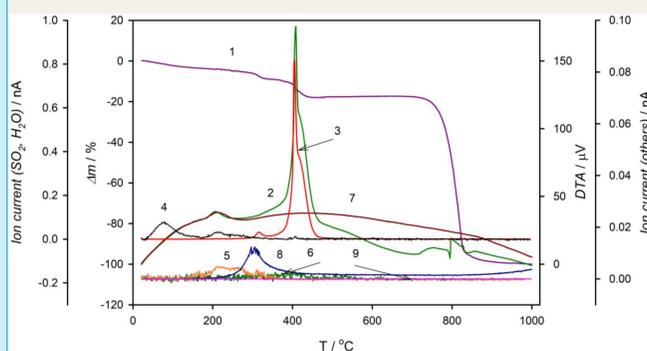
The cross-section SEM photographs show that the thickness of MoS<sub>2</sub>/Ti coatings after 5 h autoclaving approximated to 410 nm (see Insets in Fig. 2 b). The as-formed electrodes were further decorated with Pd NPs via electroless deposition method. The morphology of this deposits (Fig. 2 b, c) confirms the existence of plenty spherical-shaped Pd crystallites scattered on the MoS<sub>2</sub>/Ti surface. The deposited Pd NPs are uniformly distributed, have an average size of 24 nm and cover approximately 15.7 % of the surface's area. Direct evidence of the deposited noble metals and their loading was observed by EDX and ICP-OES analyses. From EDX investigations, the deposited crystallites could be attributed to palladium. The content of noble metals was only 1.42 atomic percentage with the 4.96  $\mu\text{g cm}^{-2}$  loading density of Pd NPs on the surface of MoS<sub>2</sub>.



**Fig. 2.** Top-side SEM photographs of nanoplatelet MoS<sub>2</sub> films fabricated on the Ti substrate by hydrothermal treatment from an aqueous solution of 5.0 mM ammonium heptamolybdate and 90 mM thiourea at 220 °C for 10 h before (a) and after chemical decoration with palladium NPs at different magnification (b, c). In (d) XRD patterns of MoS<sub>2</sub>/Ti species. Insets: top-side SEM image of leaflets edge at highest magnification and cross-section SEM image of MoS<sub>2</sub> species on Ti substrate.

## TG/DTA-MS investigations

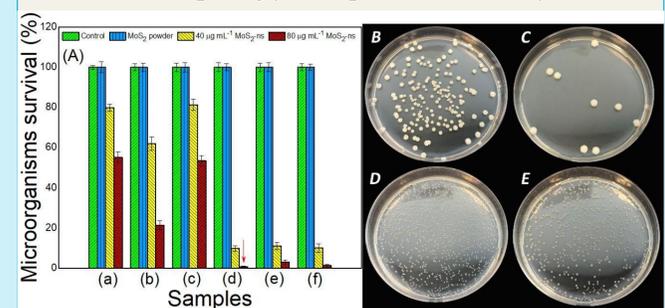
In order to determine the phase purity and composition of as-grown, petal-like MoS<sub>2</sub> powders, the TG/DTA-MS investigations were further performed herein. The evolution of sulphur under an argon atmosphere (Fig. 3 curve 8) can be linked with the presence of different chemical state of sulphur. Since the production of MoS<sub>2</sub> occurs through an intermediate MoS<sub>3</sub> stage and generation of free sulphur, this evaporation can be related to nanometric-sized sulphur residues or MoS<sub>3</sub> phase decomposition, which starts to releasing sulphur from 200 °C.



**Fig. 3.** TG/DTA-MS analysis of hydrothermally synthesized MoS<sub>2</sub> powder. TG (1) and DTA curves recorded under synthetic air (2), and argon (7) atmospheres. Curves 3, 4, 5, 6 and 8, 9 present ion current of gaseous species decomposed in synthetic air (SO<sub>2</sub>, H<sub>2</sub>O, NO/NO<sub>2</sub>, CO<sub>2</sub>) and argon (S<sub>2</sub>, CS<sub>2</sub>), respectively.

## Antimicrobial activity of MoS<sub>2</sub>-based materials

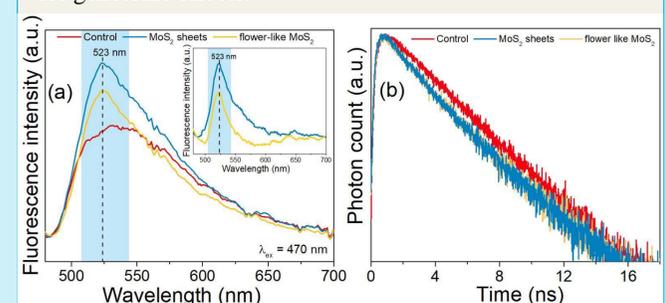
Antimicrobial activity of MoS<sub>2</sub>-ns, petal-like MoS<sub>2</sub> and MoS<sub>2</sub>-based coatings were evaluated via serial broth dilution method. The results obtained in this study clearly establishing that MoS<sub>2</sub>-ns possess the bactericidal or fungicidal potential against all tested microorganism and are summarised in Fig. 4. The microorganism survival histograms clearly show the concentration-dependent antimicrobial activity of the synthesized MoS<sub>2</sub>-ns against all tested bacteria and yeast. It is evident from the results that MoS<sub>2</sub>-ns were highly effective towards the inactivation of bacteria and surprisingly, two species of *Candida* yeast.



**Fig. 4.** Antimicrobial potency of MoS<sub>2</sub>-ns against prokaryotic *E. coli* (a), *P. aeruginosa* (b), *S. enterica* (c), *S. aureus* MRSA (d) and eukaryotic *C. parapsilosis* (e), *C. krusei* (f) microorganisms after 24 h treatment at 37 and 28 °C, respectively. Photographs showing the quantification of *C. krusei* (top row) and *E. coli* (bottom row) CFU growth inhibition induced by MoS<sub>2</sub>-ns. The leftmost Petri dishes (B, D) present control samples.

## Determination of ROS

The overproduction of ROS occurs as a consequent of MoS<sub>2</sub>-ns interaction with bacteria membrane, which turns the antioxidant defence mechanism and, thus induce oxidative stress, therefore the cells are unable to maintain the physiological redox-regulated process. Generation of ROS initiate the oxidation of proteins, induce the lipid peroxidation, cause DNA damage and could be responsible for genotoxic effects.



**Fig. 5.** Fluorescence spectra (a) and fluorescence decay kinetics (b) of DCF dye that was intracellular de-esterified by Gram-negative *Pseudomonas aeruginosa* bacteria and oxidized by ROS after exposure with MoS<sub>2</sub>-ns and flower-like MoS<sub>2</sub> particles for 2 hours at 37 °C under 150 rpm. Insets: The fluorescence spectra after subtraction of bacteria fluorescence.

The obtained photoluminescence emission peak at 523 nm wavelength is related to DCF dye fluorescence, which was obtained after exposure of *P. aeruginosa* bacteria with MoS<sub>2</sub> nano/microparticles (Fig. 5 a). It is notable that due to bacteria autofluorescence, a control sample of bacteria solution demonstrated the one broad emission peak with a maximum at 530 nm wavelength, however after subtraction of this spectrum the DCF dye emission peaks become more apparent (Fig. 5 Insets). The lifetime of *P. aeruginosa* bacteria photoluminescence exhibit two exponential decay and exceeds the average relaxation time ( $\tau_{ave}$ ) equal to 4 ns (Fig. 5 b). However, after exposure to DCF and MoS<sub>2</sub> particles, the fluorescence decay kinetics changed, resulting in shorter  $\tau_{ave}$  value of 3.4 and 3.5 ns. These results suggest that MoS<sub>2</sub> nano/microparticles induce the intracellular ROS generation, thus cause the oxidation of DCFDA dye to highly fluorescence DCF compound that was detected herein.

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